



Variation in Amino Acid Content Among Three *Aloe* Species

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Aloe species are well known as medicinal plants and are also used in various commercial products. Here, we have analyzed the amino acid contents of three *Aloe* species (*Aloe arborescens*, *A. vera* and *A. saponaria*) and have discussed the differences in amino acid levels among these three species. A total of 24 and 23 amino acids were detected in *A. vera* and *A. saponaria*, respectively, whereas only 17 amino acids were detected in *A. arborescens*. *A. vera* had high levels of ammonia, aspartic acid, glutamic acid, serine, arginine, valine, alanine, whereas *A. saponaria* contained the highest levels of ammonia, glutamic acid, aspartic acid, valine and alanine. In particular *A. vera* contained 11 times more γ -amino butyric acid than *A. arborescens* did. *A. vera* contained a greater overall concentration of amino acids than the other species did. The mean concentrations of phosphoserine and taurine were higher in *A. arborescens* than in the other *Aloe* species. In conclusion, among these *Aloe* species, *A. vera* had the highest total amino acid concentration and the greatest variation in amino acid contents.

Key Words: Amino acid, *Aloe*, *Aloe vera*, *Aloe arborescens*, *Aloe saponaria*

INTRODUCTION

Aloe is a very common genus, comprising approximately 400 species of shrubby succulent plants occurring naturally in Africa of which *Aloe vera* is the most common and well known species^{1,2}. Although *Aloe* species have been used throughout history for medicinal purposes, only a few of the 400 species are actually used traditionally as herbal medicines.

In particular, the mucilaginous gel from the parenchymatous cells in the leaf pulp of *A. vera* has been used since early times for various curative purposes³ and in recent years extensive research has been carried out on the health-promoting and medicinal properties of this plant. It has shown a variety of biological and pharmacological activities, including promotion of wound healing; hypoglycemic or antidiabetic effects and anti-inflammatory, anticancer, immunomodulatory and gastroprotective properties; as well as antifungal, antityrosinase and antioxidant activities⁴⁻⁸.

A. vera and *A. arborescens* are the most widely cultivated species of *Aloe* worldwide⁹ and is also commonly used as a medicinal plant. These species have also been reported to have anti-diabetic, antitumor, antimicrobial and antifungal activities¹⁰⁻¹³.

A. saponaria is another variety used worldwide and is also known as soap *Aloe* or African *Aloe*. This species has a lower aloin content than other *Aloe* varieties and therefore has a less bitter taste than other *Aloe* species¹⁴. The ethanol extract of *A. saponaria* displays antioxidant, antinociceptive and anti-inflammatory activities¹⁵.

In order to further understand the differences between various species of *Aloe* studies have focused on identifying the primary and secondary metabolites in the different species. However, to date, research on the amino acid content of *Aloe* species has been very limited. For instance, Sotelo *et al.*¹⁶ previously reported the analysis of amino acids in the edible flower of *A. vera* while Nicolson¹⁷ reported an analysis of the amino acids in the nectar of 16 different *Aloe* species. The aim of the present work was to compare the amino acid content of three *Aloe* species by using HPLC.

EXPERIMENTAL

The leaves of two year-old *Aloe* plants, belonging to *A. arborescens*, *A. saponaria* and *A. vera* were used for chemical analysis. The fresh samples were stored in-sealed clear polyethylene plastic bags at -80 °C until further use. The

samples were freeze dried at -80 °C in brown paper bags for 72 h and the dried samples were ground into a fine powder (40-mesh) by using a mill.

Extraction and analysis of free amino acids: The amino acid extraction and analysis methods were performed as described by Kim *et al.*¹⁸. Amino acids were extracted from freeze-dried plant tissues (1 g) with 30 mL of 70 % ethanol at 80 °C for 20 min; this step was repeated 3 times. After evaporating the ethanol, the residual water phase (30 mL) was mixed with ethyl ether (30 mL) by using a separation funnel. After separation, the water phase was freeze-dried. The extract was resuspended in 3 mL of 0.02 N HCl and filtered using a 0.45 µm syringe filter.

The amino acids in the extract were identified using an amino acid analyzer (HITACHI L-8900, Japan) equipped with an HITACHI HPLC column packed with the ion-exchange resin No. 2622 PF (4.6 mm × 60 mm) and a UV detector (VIS1, 570 nm; VIS2, 440 nm). Wako L-8500 buffer solutions PF-1, PF-2, PF-3, PF-4 and RG were used. Twenty microliters of each sample was injected and identification was performed using the ninhydrin reagent set (Wako Chemical Inc., Japan). Sample preparation and analysis were repeated three times.

RESULTS AND DISCUSSION

Amino acid contents were determined for samples of three species of *Aloe* collected from Korea and 24 different amino acids were found to be present in varying proportions. More specifically, a total of 24 and 23 amino acids were detected in the extracts from *A. vera* and *A. saponaria*, respectively, while only 17 different amino acids were identified in the extract from *A. arborescens*. The following 8 amino acids were present in both *A. vera* and *A. saponaria* but not in *A. arborescens*: threonine (Thr), isoleucine (Ile), leucine (Leu), tyrosine (Tyr), phenylalanine (Phe), β-aminoisobutyric (β-AiBA), histidine (His) and arginine (Arg).

In *A. arborescens*, phosphoserine (P-ser), carnosine (Car) and aspartic acid (Asp) were detected at much higher levels than other amino acids (Table-1). In *A. saponaria*, ammonia (NH₃), glutamic acid (Glu), Asp, valine (Val) and alanine (Ala) were particularly abundant, while the levels of the other amino acids were lower (Table-1). *A. vera* contained the highest amount of the following amino acids: NH₃, Asp, serine (Ser), Glu, Arg and Val (Table-1).

Moreover, *A. vera* contained the highest amount (90.15 µmol/g D.W.) of γ-amino butyric acid (GABA), which was approximately 11 times more than that present in *A. arborescens* (Table-1). Most of the amino acids (Ala, β-AiBA, Arg, Asp, ethanolamine, glycine [Gly], His, lysine (Lys), Ser, Thr, Tyr) were detected in *A. vera* at higher levels than in the other species (Table-1). In total, the greatest concentration of amino acids was detected in *A. vera* (2792.65 µmol/g D.W.). The amino acid, hydroxylysine (Hylys) is present only in the extract of *A. arborescens*. Carnosine levels were almost similar among the three *Aloe* species, while, Ala, proline (Pro) and Val levels were similar in *A. vera* and *A. saponaria*.

Aloe is frequently used for a variety of purposes worldwide. Compared to the other *Aloe* species, *A. vera* is the most commonly used *Aloe* species in the commercial and pharmaceutical

TABLE-1
AMINO ACID CONTENT IN THREE *Aloe* SPECIES

	<i>A. arborescens</i>	<i>A. saponaria</i>	<i>A. vera</i>
P-Ser	129.83 ± 10.01	98.74 ± 4.25	136.08 ± 1.77
Tau	32.33 ± 2.61	19.40 ± 0.86	27.49 ± 0.18
Asp	102.15 ± 0.99	219.51 ± 13.33	281.31 ± 1.55
Thr	–	37.48 ± 3.18	104.73 ± 8.23
Ser	33.97 ± 2.51	76.48 ± 9.48	268.59 ± 21.8
Glu	19.44 ± 1.03	328.51 ± 17.61	222.73 ± 10.28
Gly	5.17 ± 0.94	16.97 ± 2.64	69.55 ± 2.72
Ala	51.03 ± 1.15	122.78 ± 7.05	125.58 ± 3.77
α-ABA	7.39 ± 0.46	–	17.50 ± 0.71
Val	24.27 ± 0.12	143.43 ± 9.85	163.27 ± 3.41
Ile	–	43.38 ± 2.93	26.24 ± 1.5
Leu	–	42.56 ± 3.51	20.96 ± 0.8
Tyr	–	30.68 ± 4.58	37.90 ± 0.25
Phe	–	53.16 ± 4.36	43.58 ± 1
β-Ala	4.63 ± 0.3	37.64 ± 3.26	51.64 ± 1.38
β-AiBA	–	11.40 ± 1.36	20.51 ± 0.38
γ-ABA	7.99 ± 0.26	71.00 ± 3.88	90.15 ± 0.74
EOH NH ₂	15.20 ± 0.6	54.03 ± 3.3	64.25 ± 0.87
NH ₃	93.74 ± 4.2	440.28 ± 27.93	492.07 ± 5.63
Hylys	41.03 ± 1.51	–	–
Lys	7.55 ± 0.83	78.30 ± 5.33	114.34 ± 2.89
His	–	18.33 ± 1.6	38.91 ± 2.78
Car	111.02 ± 4.71	117.45 ± 1.96	124.20 ± 11.16
Arg	–	51.57 ± 2.74	215.45 ± 14.38
Pro	17.85 ± 7	32.26 ± 6.75	35.64 ± 0.05
Total	704.58 ± 39.23	2145.36 ± 141.71	2792.65 ± 98.23

The values represent the mean ± SD (n = 3).

P-Ser, Phosphoserine; Tau, Taurine; Asp, Aspartic acid; Thr, Threonine; Ser, Serine; Glu, Glutamic acid; Gly, Glycine; Ala, Alanine; α-ABA, α-Amino butyric acid; Val, Valine; Ile, Isoleucine; Leu, Leucine; Tyr, Tyrosine; Phe, Phenylalanine; β-Ala, β-Alanine; β-AiBA, β-amino isobutyric acid; GABA, Gamma amino butyric acid; EOH NH₂, Ethanolamine, NH₃, Ammonia; Hylys, Hydroxylysine; Lys, Lysine; His, Histidine; Car, Carnosine; Arg, Arginine; Pro, Proline.

fields. These species have been the best studied of all *Aloe* species, many researchers contend that it has more useful chemical properties than do the other species. However, other *Aloe* species have also currently attracted interest for different chemical properties and medicinal uses and indeed *A. arborescens* and *A. saponaria* have similar and in some respect, a higher content of metabolites as compared with *A. vera*. In this study, we analyzed the amino acid content of these 3 species of *Aloe* by HPLC. During amino acid analysis, a few non-protein amino acids were also detected, including γ-amino butyric acid, which is an inhibitory neurotransmitter in the central nervous system and has been implicated as a potential factor in human disease¹⁹.

Aloe vera has many more effective amino acids than other species. β-Alanine is known as the rate-limiting precursor for carnosine^{20,21}. Harris *et al.*²² discovered that the carnosine levels in skeletal muscle may increase by 60 % or more after ingesting β-Ala for 2-4 weeks. Other studies have demonstrated that β-Ala supplementation may further improve both endurance performance and reduce subjective feelings of fatigue during training^{23,24}.

Amino acids are not only the building blocks for proteins that are involved in every process that occurs in the body but also play a role in metabolism. Although hundreds of different amino acids exist in nature, only approximately 2 dozen of these are vital for human nutrition^{25,26}. The body can produce approximately half of the amino acids and must obtain the

rest from food. The different species of *Aloe* are an easily accessible and constitute an economically viable potential source of various amino acids.

REFERENCES

1. A.D. Klein and N.S. Penneys, *J. Am. Acad. Dermatol.*, **18**, 714 (1988).
2. C. Ulbricht, J. Armstrong, E. Basch, S. Basch, S. Bent, C. Dacey, S. Dalton, I. Foppa, N. Giese, P. Hammerness, C. Kirkwood, D. Sollars, C.S. Tanguay and W. Weissner, *J. Herb Pharmacother.*, **7**, 279 (2007).
3. D. Grindlay and T. Reynolds, *J. Ethnopharmacol.*, **16**, 117 (1986).
4. T. Reynolds and A.C. Dweck, *J. Ethnopharmacol.*, **68**, 3 (1999).
5. B.K. Vogler and E. Ernst, *Br. J. Gen. Pract.*, **49**, 823 (1999).
6. O.M. Grace, M.S. Simmonds, G.F. Smith and A.E. van Wyk, *J. Ethnopharmacol.*, **119**, 604 (2008).
7. A. Surjushe, R. Vasani and D.G. Saple, *Indian J. Dermatol.*, **53**, 163 (2008).
8. A. Feily and M.R. Namazi, *G. Ital. Dermatol. Venereol.*, **144**, 85 (2009).
9. Y. Gutterman and E. Chauser-Volfson, *Biochem. Syst. Ecol.*, **28**, 825 (2000).
10. M.I. Ali, N.M. Shalaby, M.H. Elgamal and A.S. Mousa, *Phytother. Res.*, **13**, 401 (1999).
11. A. Kodym and T. Bujak, *Pharmazie*, **57**, 834 (2002).
12. G.Z. Jin, H.J. Quan, J. Koyanagi, K. Takeuchi, Y. Miura, F. Komada and S. Saito, *Cancer Lett.*, **218**, 15 (2005).
13. H. Beppu, K. Shimpo, T. Chihara, T. Kaneko, I. Tamai, S. Yamaji, S. Ozaki, H. Kuzuya and S. Sonoda, *J. Ethnopharmacol.*, **103**, 468 (2006).
14. J. Li, T. Wang, Z. Shen and Z. Hu, *Acta Botan. Sin.*, **45**, 594 (2003).
15. E.A. Yoo, S.D. Kim, W.M. Lee, H.J. Park, S.K. Kim, J.Y. Cho, W. Min and M.H. Rhee, *Phytother. Res.*, **22**, 1389 (2008).
16. A. Sotelo, S. Lopez-Garcia and F. Basurto-Pena, *Plant Foods Hum. Nutr.*, **62**, 133 (2007).
17. S.W. Nicolson, *J. Chem. Ecol.*, **33**, 1707 (2007).
18. Y.K. Kim, H. Xu, N.I. Park, H.O. Boo, S.Y. Lee and S.U. Park, *J. Med. Plants Res.*, **3**, 897 (2009).
19. M. Watanabe, K. Maemura, K. Kanbara, T. Tamayama and H. Hayasaki, *Int. Rev. Cytol.*, **213**, 1 (2002).
20. W. Derave, M.S. Ozdemir, R. Harris, A. Pottier, H. Reyngoudt, K. Koppo, J.A. Wise and E. Achten, *J. Appl. Physiol.*, **103**, 1736 (2007).
21. C.A. Hill, R.C. Harris, H.J. Kim, B.D. Harris, C. Sale, L.H. Boobis, C.K. Kim and J.A. Wise, *Amino Acids*, **32**, 225 (2007).
22. R.C. Harris, M.J. Tallon, M. Dunnett, L. Boobis, J. Coakley, H.J. Kim, J.L. Fallowfield, C.A. Hill, C. Sale and J.A. Wise, *Amino Acids*, **30**, 279 (2006).
23. J.R. Hoffman, N.A. Ratamess, A.D. Faigenbaum, R. Ross, J. Kang, J.R. Stout and J.A. Wise, *Nutr. Res.*, **28**, 1 (2008).
24. A.E. Smith, A.A. Walter, J.L. Graef, K.L. Kendall, J.R. Moon, C.M. Lockwood, D.H. Fukuda, T.W. Beck, J.T. Cramer and J.R. Stout, *J. Int. Soc. Sports Nutr.*, **6**, 5 (2009).
25. G.S. Gilani, C. Xiao and N. Lee, *J. AOAC Int.*, **91**, 894 (2008).
26. D.J. Millward, D.K. Layman, D. Tomé and G. Schaafsma, *Am. J. Clin. Nutr.*, **87**, 1576 (2008).